

AOAC SMPR 2010.002

Standard Method Performance Requirements for Polymerase Chain Reaction (PCR) Methods for Detection of *Yersinia pestis* in Aerosol Collection Filters and/or Liquids

Intended Use: Laboratory use for analysis of aerosol collection filters and/or liquids

Method Developer and Independent Validation

Probability of Detection at the Acceptable Minimum Detection Level

1 Definitions

Probability of detection (POD) is the proportion of positive analytical outcomes for a qualitative method for a given matrix

at a given agent level or concentration. POD is concentration-dependent. The acceptable minimum detection level (AMDL) is the predetermined minimum level of a biological threat agent, which must be detected by the candidate method with an estimated 5% lower confidence limit on the POD of 0.95 or higher. The AMDL is dependent on the intended use.

2 Test Conditions

AMDL is 20,000 standardized *Yersinia pestis* CO-92 cells per filter; 2000 standardized cells per mL; 2000 genome equivalents per mL.

3 Acceptance Criteria

No more than one failure in 96 replicates.

Inclusivity

1 Definition

Strains or isolates or variants of the target agent(s) that the method can detect (Table 1).

Table 1. *Yersinia pestis* PCR method: Inclusivity panel

No.	Strain	Biovar	Achtman genotype	Comment	Availability ^a
YP1	CO92	O	1.ORI.c	Well-studied example of epidemic strain of pestis, recent isolate	CDC, WRAIR, USAMRIID
YP2	KIM	M	2.Med	Well-studied strain in academic circles, virulence data extensive	CDC, WRAIR, USAMRIID
YP3	Antiqua	A	1.Ant b	Ancient strain near root of tree	CDC, WRAIR, USAMRIID
YP4	Pestoides B	M	0.PE1		CDC, WRAIR, USAMRIID
YP5	Pestoides F	A	0.PE2.a	pPst negative, old strain in terms of phylogeny	CDC, WRAIR, USAMRIID
YP6	Pestoides G	A	0.PE2.b	pPst negative	CDC, WRAIR, USAMRIID
YP7	Angola	A	0.PE3	A "pestoides" in everything except name	CDC, WRAIR, USAMRIID
YP8	Nairobi	A	1.Ant a		CDC, WRAIR, USAMRIID
YP9	Harbin35	?	2 Ant	Rumored to be used or resulted from infection during experiments by Japanese BW Unit 731	CDC, WRAIR, USAMRIID
YP10	PBM19	O	1.ORI.a		CDC, WRAIR, USAMRIID
YP11	Java9	O	1.ORI	pFra negative	CDC, WRAIR, USAMRIID
YP12	A1122	O	1.ORI.a	Well-characterized U.S. isolate that is pgm- and pCD-; also has 2X large pPst plasmid	CDC, WRAIR, USAMRIID
YP13	Nicholisk 41	M	2.ANT		CDC, WRAIR, USAMRIID
YP14	Shasta		1.ORI	YE0387; SHASTA (20 OCT 54); SHASTA; HUMAN CASE; USA: CA; 1960 6LY; UCC YERS074	CDC, USAMRIID
YP15	Dodson		1.ORI	DODSON (AUG 70); HUMAN CASE: Male age 4.5 years; USA: Arizona (Tuba City); 27 JUN 67; UCC YERS073	CDC, USAMRIID
YP16	El Dorado				

^a CDC = Centers for Disease Control and Prevention; WRAIR = Walter Reed Army Institute of Research; USAMRIID = The United States Army Medical Research Institute for Infectious Diseases.

Table 2. *Yersinia pestis* PCR method: Exclusivity panel

No.	Species	Strain	Serotype	Comment	Availability ^a
YPNN1	<i>Yersinia ruckeri</i>	YERS063			USAMRIID
YPNN2	<i>Yersinia rohdei</i>	YERS062			USAMRIID
YPNN3	<i>Yersinia pseudotuberculosis</i>	PB1/+	1	Sequenced	WRAIR
YPNN4	<i>Yersinia pseudotuberculosis</i>	IP32953	1	Sequenced	WRAIR
YPNN5	<i>Yersinia pseudotuberculosis</i>	YPIII	3	Sequenced	WRAIR
YPNN6	<i>Yersinia pseudotuberculosis</i>	Pa3606	1b		WRAIR
YPNN7	<i>Yersinia pseudotuberculosis</i>	IB	1b		WRAIR
YPNN8	<i>Yersinia pseudotuberculosis</i>	EP2/+	1		WRAIR
YPNN9	<i>Yersinia pseudotuberculosis</i>	MD67	1		WRAIR
YPNN10	<i>Yersinia pseudotuberculosis</i>	1	1a		WRAIR
YPNN11	<i>Yersinia enterocolitica</i>	WA	O:8		WRAIR
YPNN12	<i>Yersinia enterocolitica</i>	8081	O:8	Sequenced	WRAIR
YPNN13	<i>Yersinia enterocolitica</i>	2516-87	O:9		WRAIR
YPNN14	<i>Yersinia kirstensenii</i>	Y231		Nonpathogenic	WRAIR
YPNN15	<i>Yersinia frederiksenii</i>	Y225		Nonpathogenic	WRAIR
YPNN16	<i>Yersinia intermedia</i>	Y228		Nonpathogenic	WRAIR
YPNN17	<i>Yersinia aldovae</i>	670-83		Nonpathogenic	WRAIR

^a WRAIR = Walter Reed Army Institute of Research; USAMRIID = The United States Army Medical Research Institute for Infectious Diseases. Version 6 approved by AOAC SPADA on January 22, 2009.

2 Test Conditions

Test inclusivity panel at AMDL.

3 Acceptance Criteria

100% expected results as defined for each strain on the panel.

Note: In the case of a negative result, retest that strain 96 times with no failures allowed to demonstrate an estimated 5% lower confidence limit on the POD of 0.95 or higher.

Exclusivity

1 Definition

Nontarget agents, which are potentially cross-reactive, that are not detected by the method (Table 2).

2 Test Conditions

Test exclusivity near neighbor panel at 10 times AMDL.

3 Acceptance Criteria

100% expected results as defined for each strain on the panel.

Note: In the case of a positive result, retest that strain 96 times with no failures allowed to demonstrate a 95% upper confidence limit on the POD of 0.05 or lower.

Environmental Interference

1 Definition

Ability of the assay to detect target organism in the presence of nontarget organisms or environmental substances and to be free of cross-reaction from environmental organisms and substances (Annex A).

2 Test Conditions

Test pooled environmental panel organisms at 10 times AMDL in the presence or absence of *Yersinia pestis* CO-92 at the AMDL. Test environmental substances as suspensions in the presence or absence of *Yersinia pestis* CO-92 at the AMDL.

3 Acceptance Criteria

100% expected results for environmental organisms (i.e., no false negatives in the presence of *Yersinia pestis* CO-92, and no false positives in the absence of *Yersinia pestis* CO-92).

Note: In the case of an unexpected result, retest individual strains 96 times with no failures allowed to demonstrate an estimated 5% lower confidence limit on the POD of 0.95 or higher. Data from environmental substances are for informational purposes only.

Collaborative Validation Study

Reproducibility

1 Definition

Precision under conditions where independent test results are obtained with the same methods on equivalent test items in different laboratories with different operators using separate instruments.

2 Test Conditions

Test *Yersinia pestis* CO-92 at AMDL and near neighbor organism at 10 times AMDL on dust-loaded filters or in dust-loaded aerosol collection liquid. At least 12 replicates per material per collaborator with 12 collaborators (four collaborators at each of three test sites).

3 Acceptance Criteria

Must produce at least 10 valid data sets. Report standard deviation of reproducibility (s_R).

POD at the AMDL Under Reproducibility Conditions (formerly termed System False-Negative Rate)

1 Definition

Rate of positive system results in a population of known positive test portions.

2 Test Conditions

Test *Yersinia pestis* CO-92 at AMDL on dust-loaded filters or in dust-loaded aerosol collection liquid. At least 12 replicates per matrix per collaborator with 12 collaborators (four collaborators at each of three test sites).

3 Acceptance Criteria

Data for target agent must demonstrate an estimated 5% lower confidence limit on the CPOD of 0.95 or higher, where CPOD is the probability of detection calculated from pooled valid collaborative data.

POD in the Absence of Analyte Under Reproducibility Conditions (formerly termed System False-Positive Rate)

1 Definition

Rate of positive system results in a population of known negative test portions.

2 Test Conditions

Test near neighbor organism at 10 times AMDL on dust-loaded filters or in dust-loaded aerosol collection liquid. At least 12 replicates per matrix per collaborator with 12 collaborators (four collaborators at each of three test sites).

3 Acceptance Criteria

Data for near neighbor must demonstrate a 95% upper confidence limit on the CPOD of 0.05 or lower, where CPOD is the probability of detection calculated from pooled valid collaborative data.

Acknowledgments

All or part of this work was funded by the Department of Homeland Security Science and Technology Directorate, award HSHQDC-08-C-00012.

AOAC SPADA approved PCR SMPRs as amended on January 22, 2009. PCR SMPRs (version 4) were revised on May 12, 2009 to reflect OMB proposal and to correct retest statistics. The final version as shown here was approved by SPADA on June 2, 2010 and contained revision to OMB requirement of 10 valid data sets for qualitative methods in the collaborative study.

ANNEX A Environmental Factors Panel

Organisms

1 Other Biothreat Agents

Bacillus anthracis Ames
Francisella tularensis subsp. *tularensis* Schu-S4
Burkholderia pseudomallei
Coxiella burnetii Nine Mile Phase I
Brucella melitensis

Ricinus communis (use ricin plant leaves as source of DNA)
Clostridium botulinum Type A

2 Cultivable Bacteria Identified as Being Present in Air and Soil

Acinetobacter lwoffii
Agrobacterium tumefaciens
Bacillus cohnii
Bacillus psychrosaccharolyticus
Bacillus benzoovorans
Bacillus megaterium
Bacillus horikoshii
Bacillus macroides
Bacteroides fragilis
Burkholderia cepacia
Burkholderia gladioli
Burkholderia stabilis
Burkholderia plantarii
Chryseobacterium indologenes
Clostridium sardiniense
Clostridium perfringens
Deinococcus radiodurans
Delftia acidovorans
Escherichia coli K12
Fusobacterium nucleatum
Lactobacillus plantarum
Moraxella nonliquefaciens
Mycobacterium smegmatis
Neisseria lactamica
Pseudomonas aeruginosa
Rhodobacter sphaeroides
Riemerella anatipestifer
Shewanella oneidensis
Staphylococcus aureus
Stenotrophomonas maltophilia
Streptococcus pneumoniae
Streptomyces coelicolor
Synechocystis
Vibrio cholerae
Legionella pneumophila
Listeria monocytogenes

3 DNA Viruses

Vaccinia virus (pox)
Adenovirus vaccine
Herpes simplex or CMV (whichever is available)

4 Microbial Eukaryotes

Freshwater Amoebae

Acanthamoeba castellanii
Naegleria fowleri

Fungi

Alternaria alternata
Aspergillus fumigatus
Aureobasidium pullulans
Cladosporium cladosporioides
Cladosporium sphaerospermum
Epicoccum nigrum
Eurotium amstelodami
Mucor racemosus

Paecilomyces variotii
Penicillium chrysogenum
Saccharomyces cerevisiae
Wallemia sebi
5 DNA from Higher Eukaryotes

Plants

Zea mays (corn)
Pollen from *Pinus* spp. (pine)
Cotton (use leaves from cotton plant as source of DNA)

Arthropods

Aedes aegypti (ATCC/CCL-125) mosquito cell line
Aedes albopictus (C6/36) mosquito
Dust mite (commercial source)
Flea (Rocky Mountain labs)
Drosophila cell line
Musca domestica (housefly; ARS, USDA, Fargo, ND)
Gypsy moth cell lines LED652Y cell line (baculovirus; Invitrogen)
Cockroach (commercial source)
Tick (*Amblyomma*)

Mammals

Mus musculus (ATCC/IB-123) mouse
Rattus norvegicus (ATCC/CRL-1896) rat
Canis familiaris (ATCC/CCL-183) dog
Felis catus (ATCC/CRL-8727) cat
Homo sapiens (HeLa) human

Avian

Chicken
6 Biological Insecticides

B. thuringiensis subsp. *israelensis*
B. thuringiensis subsp. *kurstaki*
B. thuringiensis subsp. *morrisoni*
Gypcheck for gypsy moths (*Lymantria dispar* nuclear polyhedrosis virus)
Cyd-X for codling moths (Codling moth granulosis virus)

Substances

1 Soils

Sandy
Loam
Clay
Subsoil
Silt

2 Dust

3 Powders and Chemicals

Bacillus thuringiensis powders (e.g., Dipel)
Powdered milk
Powdered infant formula (Fe fortified)
Powdered infant formula (low Fe formulation)
Powdered coffee creamer
Powdered sugar
Talcum powder
Wheat flour
Baking soda
Chalk dust
Brewer's yeast
Dry wall dust
Cornstarch
Baking powder
GABA (Gama aminobutyric acid)
L-Glutamic acid
Kaolin
Chitin
Chitosan
MgSO₄
Boric acid
Powdered toothpaste
Popcorn salt
EDTA
ZEP
Rid-X

The Environmental Factors Panel was originally approved in parts. SPADA approved the environmental organisms panel on December 13, 2007, and revised it on September 17, 2008. The soils were approved on January 22, 2009. The powders and chemicals were originally approved by SPADA on December 13, 2007, and revised on January 22, 2009. The entire Environmental Factors Panel was approved in final form as presented here on June 2, 2010.